

Synthesis of Optically Active Cyclohexenol Derivatives *via* Enzyme Catalyzed Ester Hydrolysis of 4-Acetoxy-3-methyl-2-cyclohexenone

Magnus Polla and Torbjörn Frejd*

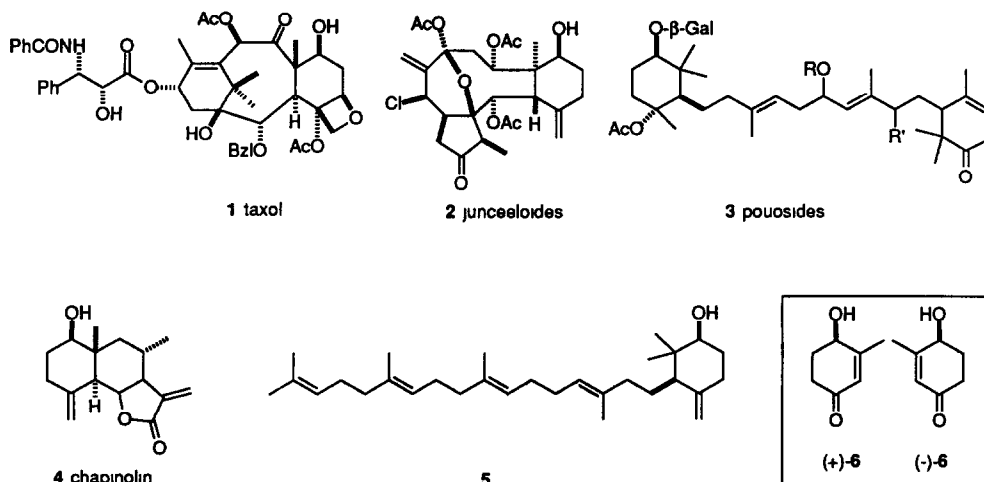
Organic Chemistry 2, Chemical Center, Lund Institute of Technology,
P O Box 124, S-221 00 Lund, Sweden

(Received in UK 22 April 1991)

Key Words Enzymatic ester hydrolyses, 1,4-additions, aldol-type reactions, absolute configuration determination, CD

Abstract The optically active cyclohexenol derivatives **9a-d**, and **10a-c** are synthesized from (-)-**6**, which is obtained by enzymatic ester hydrolysis of racemic **8**. Attempts towards the synthesis of the taxane skeleton are described

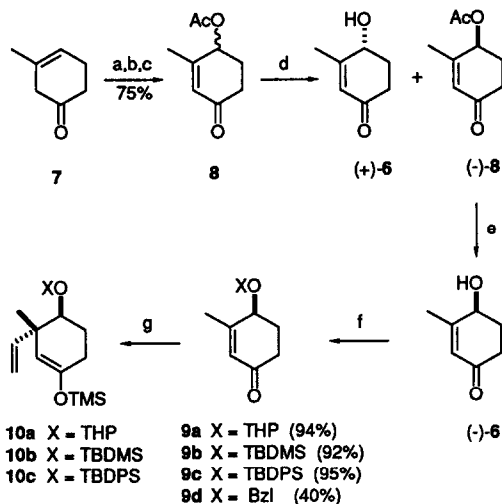
A number of natural products contain structural units related to 4-hydroxy-3-methyl-2-cyclohexenone **6**. Some examples are taxol (**1**)¹, junceeloides (**2**)², pouoside (**3**)³, chapinolin (**4**)⁴ and a recently isolated monocyclic triterpen from *Compositae* species (**5**)⁵. An efficient synthesis of both enantiomers of **6** would therefore be desirable.



We now report a short route to (+) and (-)-**6** via enzymatic asymmetric hydrolysis⁶ of the corresponding racemic acetate **8** (Scheme 1) Our attempts to use (-)-**6** for the synthesis of taxane related derivatives⁷ (Scheme 2 and 3) are also reported

RESULTS AND DISCUSSION

Epoxidation of 3-methyl-3-cyclohexenone (**7**, Scheme 2), with peracetic acid followed by triethylamine promoted isomerization of the intermediate epoxide gave the allylic alcohol **6**,⁸ which was acetylated to give **8** in 71 % overall yield from **7**. Several enzymes were then screened (0.3 M aqueous Tris-buffer at pH 7.5 and 20°C) for the hydrolysis of **8** (Table I) While a number of enzymes gave racemic material,⁹ six of them (Entries 1-6) gave an optically active product, although with modest enantiomeric excess (ee)



Scheme 1 ^a CH₃CO₂H, ^b Et₃N, ^c (Ac)₂O, Et(iPr)₂N, ^d PLE, ^e Na₂CO₃, MeOH, ^f DHP, H⁺ or TBDMSCl, imidazol or TBDPSCI, imidazol or BzlBr, Ag₂O, ^g CH₂=CHMgBr, CuBr SMe₂, TMSCl, TMEDA

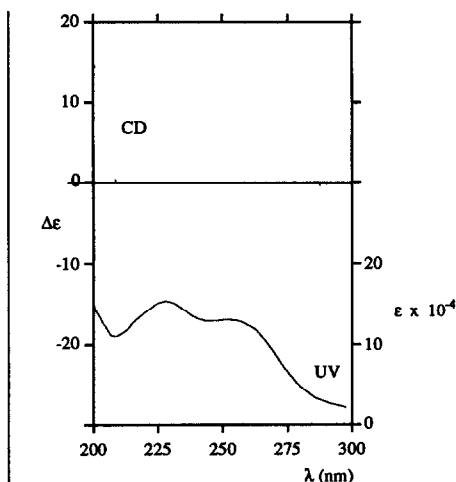


Fig 1 CD and UV spectra of the p-nitrobenzoate of (-)-**6**.

The major stereoisomer of the resulting alcohol had (R)-configuration in all cases except for the use of rabbit liver esterase (RLE) (entry 5) It is well documented that different additives or the use of mixed solvents may influence the reaction rate and the enantioselectivity of the enzymes¹⁰⁻¹⁴ The effect of the addition of some organic solvents on the reactions catalyzed by pig liver esterase (PLE) and RLE are shown in Entries 7-13 These enzymes were chosen as they most rapidly gave the highest ee's of **6** The addition of t-BuOH and diisopropyl ether to the reaction mixture containing RLE caused a dramatic decrease in reaction rate while for PLE the reaction rate was essentially unaffected However, the enantioselectivity decreased

from about 60 to 40 % ee. In the PLE-catalyzed reaction the addition of 8 % DMSO increased the ee (Entry 12), while a decrease was observed in the RLE-catalyzed reaction (Entry 9) The best result (90% ee) with PLE was obtained in 25 % DMSO (Entry 13) As reported earlier, different PLE preparations may show different degrees of selectivity^{13,14} Thus, when performing the hydrolysis in 25% DMSO using PLE from different sources (Fluka, Boehringer-Mannheim, and several batches from Sigma) the ee of (+)-6 ranged from 75 to 90 %.

Table 1 Enzymatic Hydrolysis of 8 to Give 6⁹

entry	enzyme ^a	additive	conversion (%) ^b	reaction time	ee (%) ^c of 6	config of 6
1	<i>Aspergillus niger</i> ^d	--	49	96 h	34	R
2	2212 F ^e	--	45	35 h	34	R
3	<i>Arthrobacter sp</i> ^f	--	45	2 weeks	54	R
4	Acetylcoline esterase ^g	--	40	72 h	60	R
5	Rabbit liver esterase ^g	-	40	24 h	66	S
6	Pig liver esterase ^g	--	48	2 h	63	R
7	Rabbit liver esterase	8% (iPr) ₂ O	-- ^h	--	--	--
8	Rabbit liver esterase	8% t-BuOH	-- ^h	--	--	--
9	Rabbit liver esterase	8% DMSO	47	30 h	37	S
10	Pig liver esterase	8% (iPr) ₂ O	45	2 h	42	R
11	Pig liver esterase	8% t-BuOH	45	2 h	45	R
12	Pig liver esterase	8% DMSO	41	2 h	76	R
13	Pig liver esterase	25% DMSO	45	10 h	90	R
14	Pig liver esterase	25% DMSO	62	30 h	99 ⁱ	S ⁱ

a) Lipase, unless otherwise noted. b) The reactions were monitored by GC c) The enantiomeric excess was determined from ¹H NMR spectra of the esters of (-)-menthylxyacetic acid in presence of Eu(fod)₃ d) Fluka e) Röhm

f) Sumitomo Chemical Co g) Sigma. h) Very slow reaction i) The absolute configuration and the % ee refers to that of the alcohol obtained from hydrolysis of the remaining acetate

In cases with low selectivity it is possible to obtain the remaining enantiomer in high ee by increasing the degree of conversion¹⁵ Hence, hydrolysis of the acetate remaining at 62 % conversion by the use of PLE (Entry 14), gave (-)-6 in 32% yield with >99% ee¹⁶ Enzyme catalyzed ester hydrolysis using *Arthrobacter* lipase has recently been applied to compounds structurally similar to 8, e.g. the acetate of 4-hydroxy-3-methyl-2-(2-propynyl)-2-cyclopentenone (HMPC)^{17,18} The hydrolysis of this acetate is very stereospecific and gives (+)-HMPC in 99.4% ee As seen in entry 3 the use of 8 as substrate results in a very slow reaction

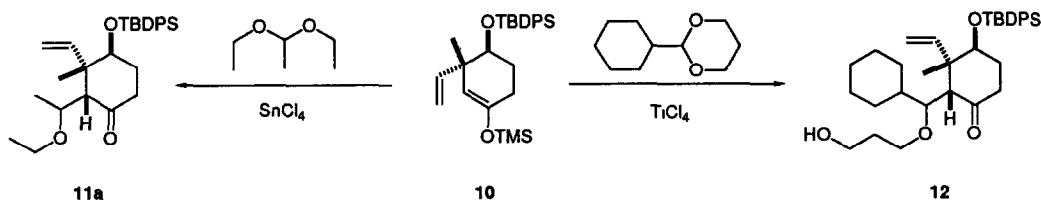
yielding (+)-**6** in only 54% ee. This low selectivity is consistent with the model of the enzyme-substrate binding site presented by Mitsuda et al. According to the model the substrate should carry a lipophilic side arm at C-2 to give the highest enantioselectivity, a structural feature that **8** is lacking.

The absolute configuration of (–)-**6** was determined by the exciton chirality method.¹⁹⁻²¹ The CD-spectrum of its *p*-nitrobenzoate (Fig. 1) shows a large negative Cotton-effect ($\Delta\epsilon_{250} -14.5$) in the region of the *p*-nitrobenzoate $\pi - \pi^*$ transition (253 nm, ϵ 13100) indicating that the compound has the (*S*)-configuration. Further support for this is obtained by comparing the experimental and the calculated CD-spectrum. The negative Cotton-effect at 250 nm is reproduced in the calculated spectrum^{22,23} although the amplitude is lower than in the experimental curve.

Selective protection of alcohol (–)-**6** as the benzyl ether **9d** without affecting the ketone proved to be an arduous task. Both basic and acidic reaction conditions gave by-products formed *via* enolisation and aromatization. The best results were obtained using Ag_2O / $\text{BzI}Br$ in DMF.²⁴ Benzyl ether **9d** was then formed in 30–40% yield in a rather capricious reaction. However, the THP, TBDMS (t-butyldimethylsilyl), and TBDPS (t-butyldiphenylsilyl) derivatives **9a-c** could be obtained in good yields, using standard procedures.

The cyclohexanol ring of all the previously mentioned natural products (**1-5**) carries a tertiary center next to the alcohol group. Thus, it is necessary to attach at C-3 of **6** a carbon chain or a grouping that could be transformed into various structures. The vinyl moiety would be one such an alternative. We therefore examined the possibility of copper mediated conjugate addition of vinylmagnesium bromide and found that the reagent formed from $\text{CuBr} \cdot \text{SMe}_2$ (1 equiv), vinylmagnesium bromide (2 equiv), (TMEDA) (2 equiv) and (TMSCl) (2.5 equiv) gave the silyl enol ethers **10a-c** in high yields²⁵ (Scheme 1). The expected anti-addition relative to the oxygen substituent at C(γ) was supported by NOESY experiments.

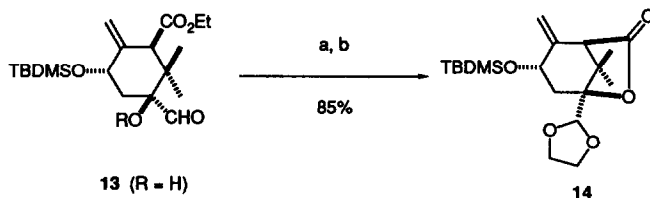
In conjunction with a project directed towards the synthesis of taxanes we envisaged that silyl enol ethers **10a-c** would be useful for the attachment of suitable A-ring units. In this respect the results of Kende et al.,²⁶ who used a Lewis acid promoted coupling of an acetal and a silyl enol ether in a synthesis of a racemic taxane skeleton, seemed encouraging. Thus, when silyl enol ether **10c** was treated with 1,1-diethoxyethane/ SnCl_4 or 2-cyclohexyl-1,3-dioxane/ TiCl_4 at -75°C , smooth reactions occurred to give mainly the axial coupling products **11** and **12**, respectively (Scheme 2).



Scheme 2

The configuration at C-2 in these derivatives is opposite to that of the corresponding position in the taxanes (C-3 in the taxane numbering system), but may be changed later in the synthesis. We therefore turned

to the synthesis of A-ring acetal **14** in order to attempt the aldol-type coupling with **10c**. Acetal **14** was synthesised from the known aldehyde **13**²⁷ by the following sequence (Scheme 3). Treatment of aldehyde **13** with 1,2-bis(trimethylsilyloxy) ethane and a catalytic amount of trimethylsilyl triflate²⁸ resulted in acetalization together with lactonization and cleavage of the TBDMS ether. Compound **14** was then obtained in 85% yield by reprotection of the hydroxyl group.



Scheme 3 ^a (TMSOCH₂)₂, TMSOTf ^b TBDMSOTf, 2,6-Lutidine

Unfortunately, no reaction was observed when acetal **14** and silyl enol ether **10c** were treated with excess SnCl₄, TiCl₄ or BF₃·OEt₂, not even at elevated temperature for several days. Equally unsuccessful was the attempted aldol reaction of the metal enolates obtained by treating silyl enol ether **10a** with MeLi (alternatively followed by ZnCl₂ to give the ZnCl-enolate or MgCl₂ to give the MgCl-enolate) with aldehyde **13** (R = Li or TMS)²⁹.

EXPERIMENTAL.

All liquid chromatography separations were performed using Merck SiO₂ 60 (0.040–0.063 mm) silica gel. TLC analyses were done on Merck SiO₂ 60 F₂₅₄ precoated aluminum sheets and the spots were visualized with UV light or by charring with 5% molybdatophosphoric acid in ethanol. NMR-spectra were recorded at 23°C with a Varian XL-300 spectrometer operating at 300 MHz proton frequency (software version 6.2) using CDCl₃ as solvent and CHCl₃ as internal standard (δ 7.26 ppm as compared to TMS). Optical rotations were measured with a Perkin-Elmer 141 polarimeter. Mass spectra were recorded on a Finnigan 4021 spectrometer (electron impact mode) and a Jeol JMS-SX 102 for the high resolution mass spectra. CD spectra were recorded on a Jasco J-41A spectropolarimeter and UV spectra were obtained with a Varian Carry 2290 Spectrophotometer. GC analyses were carried out on a Varian 3700 gas chromatograph equipped with a RSL-300 capillary column at 130°C. Vinylmagnesium bromide in THF was purchased from Alfa Ventron. Magnesium sulfate was used as drying reagent for organic extracts.

4-Hydroxy-3-methyl-2-cyclohexenone (6) Peracetic acid³⁰ (10.0 mL, 1 M, 10.0 mmol) saturated with NaOAc, was added dropwise over 30 min to a stirred suspension of Na₂CO₃ (10.0 g, 94.3 mmol) and 7³¹ (0.87 g, 7.9 mmol) in CH₂Cl₂ (125 mL). After 120 min of additional stirring, the reaction mixture was washed with saturated aqueous NaHCO₃ and dried. Triethylamine (2.2 mL, 16 mmol) was added and the

resulting solution was stirred for 150 min. The reaction mixture was filtered, washed with brine, dried and concentrated under reduced pressure. Caution! Peracids may form explosive peroxides. Chromatography of the residue (heptane-EtOAc, 1:1) gave **6** (0.75 g, 75 %) as a pale yellow oil. $^1\text{H NMR}$ δ 5.85 (m, 1 H, 2-H), 4.33-4.42 (m, 1 H, 4-H), 2.52-2.62 (m, 1 H, 6-H), 2.24-2.41 (m, 2 H, 5-H, 6-H), 2.05 (m, 3 H, CH_3), 1.95-2.04 (m, 1 H, 5-H). Anal. Calcd for $\text{C}_7\text{H}_{10}\text{O}_2$: C, 66.65, H, 7.99. Found: C, 66.54, H, 8.02. EIMS m/e 126 (M^+).

4-Acetoxy-3-methyl-2-cyclohexenone (8) Ethyldiisopropylamine (3.3 mL, 19 mmol) was added to a solution of **6** (1.0 g, 8.0 mmol), acetic anhydride (2.0 mL, 20 mmol) and 4-pyrrolidinopyridine (0.10 g, 0.7 mmol) in CH_2Cl_2 (20 mL). After stirring for 120 min, the reaction mixture was diluted with EtOAc and washed successively with saturated aqueous NaHCO_3 and water. The organic phase was dried and concentrated under reduced pressure. Chromatography of the residue (heptane-EtOAc, 3:1) yielded **8** (1.25 g, 93.0 %) as a pale yellow oil. $^1\text{H NMR}$ δ 5.94 (m, 1 H, 2-H), 5.52-5.60 (m, 1 H, 4-H), 2.50-2.60 (m, 1 H, 6-H), 2.22-2.45 (m, 2 H, 5-H, 6-H), 2.14 (s, 3 H, COCH_3), 2.01-2.14 (m, 1 H, 5-H), 1.94 (t, 3 H, CH_3). HRMS Calcd for $\text{C}_9\text{H}_{12}\text{O}_3$: 168.0783, Found: 168.0758. The acetate **8** was not stable and was used immediately in the following step.

Screening of enzymes The appropriate enzyme (1 mg) followed by compound **8** (50 mg, 0.31 mmol) was added to a stirred solution of organic solvent (x mL) in 0.3 M aqueous tris(hydroxymethyl)amino-methane hydrochloride (Tris-HCl) ($20-x$ mL, pH 7.5) at 20°C . The reaction was monitored by GC analysis and terminated at ca 45% conversion. The reaction mixture was then extracted with EtOAc (3x40 mL) and the combined organic layers were washed with brine, dried and concentrated. Chromatography of the residue (heptane-EtOAc, 3:1) gave alcohol **6** as a pale yellow oil, which was esterified with (-)-menthylxyacetic acid.³² The ee was determined by NMR analysis of the ester in the presence of $\text{Eu}(\text{fod})_3$.

(S)-4-Acetoxy-3-methyl-2-cyclohexenone ((-)-8) PLE (~25 mg) followed by compound **8** (0.95 g, 7.6 mmol) was added to a solution of DMSO (40 mL, 25%) in 0.3 M aqueous Tris-HCl (120 mL, pH 7.0) at 20°C . The slowly decreasing pH was kept constant throughout the reaction by continuous addition of 0.1 M aqueous NaOH from a Radiometer pH-stat. After stirring for 30 h (62% conversion) the reaction mixture was extracted with EtOAc (3 x 250 mL) and the combined organic layers were washed with brine, dried and concentrated. Chromatography of the residue (heptane-EtOAc, 3:1) gave (-)-**8** (0.34 g, 36%) as a pale yellow oil. $[\alpha]_{\text{D}}^{25} - 35.1^\circ$ (c 0.61, CDCl_3), >99% ee. The spectral data are identical with those of racemic **8**.

(S)-4-Hydroxy-3-methyl-2-cyclohexenone ((-)-6) Na_2CO_3 (2.0 g, 19 mmol) was added to a solution of (-)-**8** (0.34 g, 2.0 mmol) in CH_3OH (15 mL). After stirring for 120 min, the reaction mixture was filtered and concentrated under reduced pressure. Flash chromatography of the residue (heptane-EtOAc, 1:3) gave (-)-**6** (0.23 g, 90%). $[\alpha]_{\text{D}}^{25} - 48.8^\circ$ (c 0.98, CDCl_3). The spectral data are identical with those of racemic **6**.

(4S)-3-Methyl-4-[(tetrahydropyranyl)oxy]-2-cyclohexenone (9a). Compound **9a** was prepared from (-)-**6** (0.50 g, 4.0 mmol), DHP (0.60 mL, 6.6 mmol) and pyridinium tosylate (0.10 g, 0.40 mmol) in CH_2Cl_2 (20 mL).³³ Chromatography (heptane-EtOAc, 5:1) gave **9a** (0.78 g, 94%) as a clear oil $[\alpha]_{\text{D}}^{25} - 36.3^\circ$ (c 0.81, CDCl_3) $^1\text{H NMR}$ δ 5.83, 5.89 (2 m, 1 H, 2-H), 4.75-4.82 (m, 1 H, THP), 4.24, 4.39 (2 m, 1 H, 4-H), 3.57, 3.92 (2 m, 2 H, THP), 2.50-2.63 (m, 1 H, 6-H), 2.09-2.40 (m, 3 H, 5-H, 6-H), 1.98, 2.08 (2 m, 3 H, CH_3), 1.50-1.90 (m, 6 H, THP) Anal Calcd for $\text{C}_{12}\text{H}_{18}\text{O}_3$ C, 68.55, H, 8.63 Found C, 68.39, H, 8.72 EIMS m/e 210 (M^+)

(S)-4-[(t-Butyldimethylsilyl)oxy]-3-methyl-2-cyclohexenone (9b). Compound **9b** was prepared from (-)-**6** (0.10 g, 0.79 mmol), TBDMSCl (0.092 g, 1.3 mmol) and imidazol (0.17 g, 2.5 mmol) in DMF (1.0 mL).³⁴ Chromatography (heptane-EtOAc, 10:1) gave **9b** (92%) as an oil $[\alpha]_{\text{D}}^{25} - 32.1^\circ$ (c 1.16, CDCl_3) $^1\text{H NMR}$ δ 5.82, (q, 1 H, 2-H), 4.35 (dd, 1 H, 2-H), 2.50-2.60 (m, 1 H, 6-H), 2.25-2.37 (m, 1 H, 6-H), 2.11-2.22 (m, 1 H, 5-H), 1.91-2.05 (m, 1 H, 5-H), 1.98 (m, 3 H, CH_3), 0.92 (s, 9 H, t-Bu), 0.13 (2 s, 6 H, SiMe_3) Anal Calcd for $\text{C}_{13}\text{H}_{24}\text{O}_2\text{Si}$ C, 64.94, H, 10.06 Found C, 64.58, H, 10.29 EIMS m/e 183 ($\text{M}^+ - \text{t-Bu}$)

(S)-4-[(t-Butyldiphenylsilyl)oxy]-3-methyl-2-cyclohexenone (9c). Compound **9c** was prepared from (-)-**6** (0.10 g, 0.79 mmol), TBDPSCl (0.23 mL, 1.5 mmol), imidazol (0.20 g, 3.0 mmol) in DMF (1.0 mL).³⁴ Chromatography (heptane-EtOAc, 10:1) gave **9c** (95%) $[\alpha]_{\text{D}}^{25} + 4.7^\circ$ (c 1.03, CDCl_3) $^1\text{H NMR}$ δ 7.73, 7.48 (2m, 10 H, phenyl), 5.80 (m, 1 H, 2-H), 4.35 (m, 1 H, 4-H), 2.45-2.56 (m, 1 H, 6-H), 1.87-2.17 (m, 3 H, 5-H, 6-H), 1.95 (m, 3H, CH_3), 1.09 (s, 9 H, t-Bu) Anal Calcd for $\text{C}_{23}\text{H}_{28}\text{O}_2\text{Si}$ C, 75.78, H, 7.74 Found C, 75.7, H, 8.0 EIMS m/e 307 ($\text{M}^+ - \text{t-Bu}$)

4-[(Benzyl)oxy]-3-methyl-2-cyclohexenone (9d). Compound **9d** was prepared from **6** (0.10 g, 0.79 mmol), BzlBr (0.50 g, 2.9 mmol) and Ag_2O (1.0 g, 4.3 mmol) in DMF (3 mL).²⁴ Chromatography (heptane-EtOAc, 5:1) gave **9d** (30-40%) $^1\text{H NMR}$ δ 7.28-7.40 (m, 5 H, phenyl), 5.87 (s, 1 H, 2-H), 4.65 (dd, 2 H, $\text{CH}_2\text{C}_6\text{H}_5$), 4.07 (m, 1 H, 4-H), 2.55-2.65 (m, 1 H, 6-H), 2.21-2.36 (m, 2H, 5-H, 6-H), 2.04-2.17 (m, 2 H, 5-H), 2.02 (m, 3 H, CH_3) Anal Calcd for $\text{C}_{14}\text{H}_{16}\text{O}_2\text{Si}$ C, 77.75, H, 7.46 Found C, 77.8, H, 7.4

(3S,4S)-3-Methyl-4-[(tetrahydropyranyl)oxy]-1-[(trimethylsilyl)oxy]-3-vinylcyclohexene (10a). Vinylmagnesium bromide (1.92 mL of a 1.0 M solution in THF, 1.92 mmol) was added dropwise during 20 min to $\text{CuBr} \cdot \text{SMe}_2$ (196 mg, 0.960 mmol) in THF (4 mL) at -75°C under nitrogen. The mixture was stirred at -75°C for 20 min. Then TMEDA (0.28 mL, 1.9 mmol) was added, followed by TMSCl (0.30 mL, 2.4 mmol) and a solution of **9a** (0.10 g, 0.48 mmol) in THF (0.5 mL). The reaction mixture was stirred at -75°C for 180 min whereafter the cooling bath was allowed to warm slowly to room temperature. After stirring for 16 h, the reaction mixture was diluted with hexane, washed successively with water and saturated aqueous NaHCO_3 , dried and concentrated under reduced pressure to give crude **10a** (0.150 g, 100%). A small amount

of the compound was purified by short column flash chromatography (hexane) for characterization $^1\text{H NMR}$ δ 5.77-5.93 (m, 1 H, vinyl), 4.95-5.06 (m, 2 H, vinyl), 4.64, 4.77 (2 m, 1 H, THP), 4.53, 4.57 (2 m, 1 H, 2-H), 3.87-3.97 (m, 1 H, THP), 3.44-3.62 (2 m, 3 H, 4-H, THP), 1.50-2.28 (m, 10 H), 1.08, 1.16 (2 s, 3 H, CH_3), 0.20 (s, 9 H, TMS) Anal. Calcd for $\text{C}_{17}\text{H}_{30}\text{O}_3\text{Si}$ C, 65.76; H, 9.74 Found C, 65.82, H, 9.78. EIMS m/e 311 (M^+)

(3S,4S)-4-[(*t*-Butyldimethylsilyl)oxy]-3-methyl-1-[(trimethylsilyl)oxy]-3-vinylcyclohexene (10b) was prepared in a quantitative crude yield as above using 9b $^1\text{H NMR}$ data for crude 10b δ 5.80 (dd, 1 H, vinyl), 5.00 (dd, 2 H, vinyl), 4.95 (dd, 1 H, vinyl), 3.54 (dd, 1 H, 4-H), 2.07-2.19 (m, 1 H, 6-H), 1.89-2.01 (m, 1 H, 6-H), 1.62-1.75 (m, 2 H, 5-H), 1.04 (s, 3 H, CH_3), 0.89 (s, 9 H, *t*-Bu), 0.19 (s, 9 H, TMS), 0.04 (2s, 6 H, $\text{Si}(\text{CH}_3)_2$) HRMS calcd for $\text{C}_{18}\text{H}_{36}\text{O}_2\text{Si}_2$ 340.2244 Found 340.2289.

The crude enol ether could not be purified further but was instead hydrolyzed to give the corresponding ketone for characterization. Thus, crude 10b (0.16 g, 0.48 mmol) was added to a mixture of silica gel (ca 10 mg) and CH_3OH /water (10/1, 1 mL). After stirring for 16 h the reaction mixture was diluted with EtOAc and washed with water. The organic phase was dried and concentrated at reduced pressure. Chromatography of the residue (heptane-EtOAc, 3/1) gave (3S,4S)-4-[(*t*-butyldimethylsilyl)oxy]-3-methyl-3-vinylcyclohexanone (0.10 g, 81%) as an oil. $^1\text{H NMR}$ δ 5.67 (dd, 1 H, vinyl), 5.02 (2 dd, 2 H, vinyl), 3.69 (m, 1 H, 4-H), 2.44 (dd, 2 H, 2-H), 2.53 (m, 1 H, 6-H), 2.15 (m, 1 H, 6-H), 1.93-2.06 (m, 1 H, 5-H), 1.78-1.88 (m, 1 H, 5-H), 1.07 (s, 3 H, CH_3), 0.93 (s, 9 H, *t*-Bu), 0.12 (2 s, 6 H, $\text{Si}(\text{CH}_3)_2$) Anal. Calcd for $\text{C}_{15}\text{H}_{28}\text{O}_2\text{Si}$ C, 67.10, H, 10.51 Found C, 67.30; H, 10.27 EIMS m/e 211 ($\text{M}^+ - \text{t-Bu}$)

(3S,4S)-4-[(*t*-butyldiphenylsilyl)oxy]-3-Methyl-1-[(trimethylsilyl)oxy]-3-vinylcyclohexene (10c) was prepared in a quantitative crude yield as above using 9c $^1\text{H NMR}$ data for crude 10c δ 7.69, 7.39 (2 m, 10 H, phenyl), 5.78 (dd, 1 H, vinyl), 4.97 (dd, 1 H, vinyl), 4.92 (dd, 1 H, vinyl), 4.52 (s (broad), 1 H, 2-H), 3.70 (dd, 1 H, 4-H), 1.92-2.03 (m, 1 H, 6-H), 1.47-1.83 (m, 3 H, 5-H, 6-H), 1.14 (s, 3 H, CH_3), 1.05 (s, 9 H, *t*Bu), 0.17 (s, 9 H, TMS) HRMS calcd for $\text{C}_{28}\text{H}_{40}\text{O}_2\text{Si}_2$ 464.2556 Found 464.2574

The crude enol ether could not be purified further but was instead hydrolyzed as above to give the corresponding ketone for characterization. Thus, crude 10c (0.22 g, 0.48 mmol) gave (3S,4S)-4-[(*t*-butyldiphenylsilyl)oxy]-3-methyl-3-vinylcyclohexanone (0.17 g, 90%) after hydrolysis, work-up and chromatography (hexane-EtOAc, 10/1) $^1\text{H NMR}$ δ 7.70, 7.41 (2 m, 10 H, phenyl), 5.61 (dd, 1 H, vinyl), 4.96 (2 dd, 2 H, vinyl), 3.85 (dd, 1 H, 4-H), 2.47 (dd, 2 H, 2-H), 2.41-2.53 (m, 1 H, 6-H), 1.99-2.09 (m, 1 H, 6-H), 1.65-1.83 (m, 2 H, 5-H), 1.11 (s, 9 H, *t*-Bu), 1.06 (s, 3 H, CH_3) Anal. Calcd for $\text{C}_{25}\text{H}_{33}\text{O}_2\text{Si}$ C, 76.48, H, 8.22 Found C, 76.37, H, 8.35 EIMS m/e 307 ($\text{M}^+ - \text{t-Bu}$)

(2S, 3S, 4S)-4-[(t-butylidiphenylsilyloxy)-2-(1-ethoxyethyl)-3-Methyl-3-vinyl cyclohexanone (11a) and (2R, 3S, 4S)-4-[(t-butylidiphenylsilyloxy)-2-(1-ethoxyethyl)-3-Methyl-3-vinylcyclohexanone (11b). SnCl₄ (0.12 mL of a 1.0 M solution in CH₂Cl₂, 0.12 mmol) was added dropwise to a solution of compound 10c (56 mg, 0.12 mmol) and 1,1-diethoxyethane (16 μL, 0.12 mmol) in CH₂Cl₂ (1.0 mL) at -75°C under nitrogen. After stirring for 90 min, the reaction mixture was diluted with diethyl ether and washed with saturated aqueous NaHCO₃. The organic phase was dried and concentrated at reduced pressure. Chromatography of the residue (heptane-EtOAc, 10/1) gave 11a (39 mg, 71 %) and 11b (8 mg, 14 %).

11a ¹H NMR δ 7.72, 7.38 (2 m, 10 H, phenyl), 5.76 (dd, 1 H, vinyl), 4.99 (dd, 1 H, vinyl), 4.93 (dd, 1 H, vinyl), 3.78 (dd, 1 H, 4-H), 3.58-3.66 (m, 1 H), 3.47-3.57 (m, 1 H), 3.20-3.30 (m, 1 H), 2.95 (dd, 1 H, 2-H), 2.54-2.64 (m, 1 H, 6-H), 2.05-2.14 (m, 1 H, 6-H), 1.72-1.91 (m, 2 H, 5-H), 1.24 (s, 3 H, CH₃), 1.18 (d, 3 H, CH₃), 1.11 (t, 3 H, CH₃), 1.10 (s, 9 H, t-Bu)

11b ¹H NMR δ 7.80, 7.42 (2 m, 10 H, phenyl), 5.69 (dd, 1 H, vinyl), 4.98 (dd, 1 H, vinyl), 4.96 (dd, 1 H, vinyl), 3.87 (dd, 1 H, 4-H), 3.65-3.74 (m, 1 H), 3.43-3.55 (m, 1 H), 3.17-3.27 (m, 1 H), 2.47 (d, 1 H, 2-H), 2.02-2.22 (m, 2 H, 6-H), 1.73-1.84 (m, 2 H, 5-H), 1.14 (s, 3 H, CH₃), 1.06-1.16 (m, 6 H, 2 CH₃), 1.03 (s, 9 H, t-Bu) Anal. Calcd for C₂₉H₄₀O₃Si C, 74.95, H, 8.68 Found. C, 74.58, H, 8.60 CIMS m/e 464 (M⁺)

(2S, 3S, 4S)-4-[(t-butylidiphenylsilyloxy)-2-(cyclohexyl-[(3-hydroxypropyl)oxy] methyl)-3-Methyl-3-vinylcyclohexanone (12). TiCl₄ (82 μL of a 1.0 M solution in CH₂Cl₂, 82 μmol) was added dropwise to a solution of compound 10c (38 mg, 82 μmol) and 2-cyclohexyl-1,3-dioxane (14 mg, 82 μmol) in CH₂Cl₂ (1.0 mL) at -75°C under nitrogen. After stirring for 60 min, the reaction mixture was diluted with diethyl ether and washed with saturated aqueous NaHCO₃. The organic phase was dried and concentrated at reduced pressure. Chromatography of the residue (hexane-EtOAc, 5/1) gave 12 (28 mg, 60 %) ¹H NMR δ 7.70, 7.41 (2 m, 10 H, phenyl), 5.71 (dd, 1 H, vinyl), 4.93 (dd, 1 H, vinyl), 4.83 (dd, 1 H, vinyl), 3.85 (dd, 1 H, 4-H), 3.54-3.79 (m, 5 H), 3.34 (d, 1 H, 2-H), 2.61-2.72 (m, 1 H, 6-H), 1.10-2.35 (m, 17 H), 1.27 (s, 3 H, CH₃), 1.16 (s, 9 H, t-Bu) Anal. Calcd for C₃₅H₅₀O₄Si C, 74.68, H, 8.95 Found. C, 74.34, H, 9.38 CIMS m/e 563 (M⁺+H)

(1S, 3S, 5S)-1-[(t-Butyldimethylsilyloxy)-5-(1, 3-dioxo-2-cyclopentyl)-2-methylene-4, 4-dimethyl-3, 5-cyclohexanecarbolactone (14) Bis(trimethylsilyloxy)ethane (22 μL, 88 μmol) and a solution of compound 13 (R = H)²⁷ (30 mg, 81 μmol) in CH₂Cl₂ (0.5 mL) was added successively to a solution of trimethylsilyltriflate (1.6 μL, 9 μmol) in CH₂Cl₂ (3 mL) at -75°C under nitrogen. The reaction mixture was stirred at -75°C for 300 min, whereafter the cooling bath was allowed to warm slowly to room temperature. After stirring for 48 h, the reaction mixture was diluted with diethyl ether and washed with saturated aqueous NaHCO₃. The organic phase was dried and concentrated. Short column flash chromatography of the residue (hexane-EtOAc, 1/1) gave (1S, 3S, 5S)-5-(1, 3-dioxo-2-cyclopentyl)-1-hydroxy-2-methylene-4, 4-dimethyl-3, 5-cyclohexanecarbolactone (18 mg, 87 %) ¹H NMR δ 5.32 (d, 1 H, methylene), 5.05 (d, 1 H, methylene), 5.03 (s, 1 H, acetal), 4.43 (m, 1 H, 3-H), 3.87-4.03 (m, 4 H, acetal), 2.85 (s, 1 H, 5-H), 2.64 (dd, 1 H, 2-H),

2.02 (d, 1H, OH), 1.68 (dd, 1H, 2-H), 1.20 (s, 3H, CH₃), 1.11 (s, 3H, CH₃) CIMS m/e 255 (M⁺+H)
 t-Butyldimethylsilyltriflate (24 μL, 0.10 mmol) was added to a solution of the lactone (18 mg, 68 μmol) and 2,6-lutidine (16 μL, 0.14 mmol) in CH₂Cl₂ (0.5 mL). After stirring for 60 min, the reaction mixture was diluted with diethyl ether and washed successively with 1 M aqueous HCl and saturated aqueous NaHCO₃. The organic phase was dried and concentrated under reduced pressure. Short column flash chromatography of the residue (hexane-EtOAc, 10:1) gave **14** (25 mg, 98%) as a white solid. Mp 113°C, [α]_D²⁵ 119.8° (c 0.28, CDCl₃), ¹H NMR δ 5.27 (d, 1H, =CH₂), 5.01 (s, 1H, acetal), 4.96 (d, 1H, =CH₂), 4.34 (m, 1H, 3-H), 3.85-4.04 (m, 4H, acetal), 2.85 (s, 1H, 5-H), 2.46 (dd, 1H, 2-H), 1.66 (dd, 1H, 2-H), 1.09 (s, 3H, CH₃), 1.19 (s, 3H, CH₃), 0.92 (s, 9H, t-Bu), 0.08 (s, 6H, Si(CH₃)₂) Anal. Calcd for C₁₉H₃₂O₅Si. C, 61.92, H, 8.75. Found C, 61.65, H, 8.63. CIMS m/e 369 (M⁺+H)

Attempted reaction between acetal 14 and silyl enol ether 10c TiCl₄ (27 μL of a 0.8 M solution in CH₂Cl₂, 22 μmol) was added to a solution of **14** (5 mg, 14 μmol) and **10c** (10 mg, 21 μmol) in CH₂Cl₂ (0.5 mL) at -75°C under argon. The reaction mixture was stirred for 120 min followed by 48 h additional stirring at room temperature. The reaction was also performed with the following modifications: a) SnCl₄ or BF₃·OEt₂ were used instead of TiCl₄, b) 2 equiv of TiCl₄ was used instead of 1.0 equiv, c) the reaction mixture was heated at reflux for 24 h. In no case was any coupling product observed.

Attempted aldol reaction between aldehyde 13 and metal enolates of 10a The metal enolates were prepared as follows. MeLi (38 μL of a 1.6 M solution in diethyl ether, 60 μmol) was added to a solution of **10a** (67 mg, 60 μmol) in diethyl ether (0.3 mL). The reaction mixture was stirred for 30 min at room temperature to give the lithium enolate of **10a**. Subsequent addition of anhydrous ZnCl₂ (8.4 mg, 60 μmol) or MgCl₂ (6.6 mg, 60 μmol) followed by 15 min of additional stirring gave the ZnCl- and MgCl-enolates, respectively.

A solution of **13** (R = H, 20 mg, 54 μmol) in diethyl ether (0.2 mL) was added to a solution of LDA (54 μmol) in diethyl ether (0.1 mL). The reaction mixture was cooled to -50°C and the appropriate metal enolate solution was added dropwise during 5 min. Stirring was continued for 240 min at -50°C followed by 24 h at room temperature. Saturated aqueous NH₄Cl or 1 M aqueous HCl was then added, the mixture was extracted with diethyl ether and the collected organic phases were dried, concentrated and analyzed by NMR. The reaction was also performed with the following modifications: a) THF was used as solvent instead of diethyl ether, b) compound **13** (R = TMS) was used instead of aldehyde **13** (R = Li), c) the reaction was quenched at -50°C instead of room temperature. No aldol product could be detected in any of these experiments.

Acknowledgment We thank professor Jan Sandstrom for various discussions. We are also grateful to Sumitomo Chemical Co., Ltd. for a generous gift of *Arthrobacter* lipase and to Dr. S. Nabeshima at Sumitomo Chemical Co., Ltd. for a preprint of their work on this enzyme. Financial support from The Swedish Natural Science Research Council is gratefully acknowledged.

REFERENCES AND NOTES

- 1 Wani, M.C , Taylor, M L , Wall, M E , Coggon, P , McPhail, A T *J Am Chem Soc* **1971**, *93*, 2325
- 2 Shun, J., Park, M , Fenical, W *Tetrahedron* **1989**, *45*, 1633.
- 3 Ksebati, M B , Schmitz, F J , Gunasekara, S P *J Org Chem* **1988**, *53*, 3917
- 4 Vargas, D , Urbatsch, L E , Fischer, N H *Phytochemistry*, **1988**, *27*, 1413
5. Barrero, A F , Alvarez-Manzaneda R , E J , Alvarez-Manzaneda R , R *Tetrahedron Lett* **1989**, *30*, 3351
- 6 For leading references to enzymes in organic synthesis see Jones, J B *Tetrahedron* **1986**, *42*, 3351, 'Enzymes In Organic Synthesis ', Ciba Foundation Symposium 111, Porter, R , Clark, S , eds , Pitman, London, **1985**
- 7 Synthesis of compounds similar to **10** but lacking the oxygen functionality at C-4 has been reported Shubuya, H , Tsuji, S , Yamamoto, Y , Miura, H , Kitagawa, I *Chem Pharm Bull* **1984**, *32*, 3417
- 8 The synthesis of racemic **6** was reported without details by Ruden, R A , Litterer, W E *Tetrahedron Lett* **1975**, 2043
- 9 The following enzymes were also tested but gave only racemic products Porcine pancrease (Sigma), *Candida cylindracea* (Sigma), *Pseudomonas fluorescens* (Fluka), *Rhizopus delemar* (Fluka), *Rhizopus niveus* (Fluka), *Mucor javanicus* (Fluka), *Penicillium roqueforti* (Fluka), *Candida lipolytica* (Fluka), 2212 E (Rohm), *Rhizopus arrhizus* (Fluka)
- 10 Bjorkling, F , Boutelje, J , Gatlenbeck, S , Hult, K , Norin, T , Szmulik, P *Bioorganic Chemistry* **1986**, *14*, 176
- 11 Bjorkling, F , Boutelje, J , Hjalmarsson, M ; Hult, K , Norin, T *J C S Chem Comm* **1987**, 1041
- 12 Guanti, G , Banfi, L , Narisano, E *Tetrahedron Lett* **1989**, *30*, 2697
- 13 Guanti, G , Banfi, L , Narisano, E , Riva, R , Thea, S *Tetrahedron Lett* **1986**, *27*, 4639
- 14 Lam, L K P , Hui, R A H F , Jones, J B *J Org Chem* **1986**, *51*, 2047
- 15 For a quantitative treatment of biochemical kinetic resolution, see Chen, C -S , Fujimoto, Y , Girdukas, G , Sih, C J *J Amer Chem Soc* **1982**, *104*, 7294
- 16 (S)-4-Hydroxy-2-cyclohexenone, structurally similar to (-)-**6**, was recently synthesized via two routes Audia, J E , Boisvert, L , Patten, A D , Villalobos, A , Danishefsky, S J *J Org Chem* **1989**, *54*, 3738, Carreno, M C , Ruano, J L G , Garrido, M , Ruiz, M P , Solladie, G *Tetrahedron Lett* **1990**, *31*, 6653
- 17 Mitsuda, S , Nabeshima, S , Hirohara, H *Appl Microbiol Biotechnol* **1989**, *34*, in press
- 18 Mitsuda, S , Umemura, T , Hirohara, H *Appl Microbiol Biotechnol* **1988**, *29*, 310
- 19 Harada, N , Nakanishi, K *Acc Chem Res* **1972**, *5*, 257

- 20 Koreeda, M , Harada, N , Nakanishi, K *J Amer Chem Soc* **1974**, *96*, 266.
21. Harada, N , Nakanishi, K ` *Circular Dichroic Spectroscopy - Exciton Coupling In Organic Stereochemistry* ` University Science Books, Mill Valley, California, **1983**
- 22 Minimum energy conformations of the p-nitrobenzoate of (-)-**6** were calculated with the MM2(85) program: Burkert, U ; Allinger, N. L *Molecular Mechanics*, American Chemical Society, Washington D. C , U S A , 1982 The program is available through QCPE, Department of Chemistry, Indiana University, Bloomington, Indiana 47405, U S A , for academic users, and through Molecular Design Ltd , 2132 Farallon drive, San Leandro, California 94577, U S A , for non-academic users
- 23 Guimon, C , Gonbeau, D , Pfister-Guillouzo, G *Tetrahedron* **1973**, *29*, 3399 Bayley, P M ; Nielsen, E B , Schellman, J A *J Phys Chem* **1969**, *73*, 228 Rizzo, V , Schellman, J A *Biopolymers* **1973**, *23*, 435
- 24 Kuhn, R., Low, I , Trischman, H *Chem Ber* **1957**, 203
- 25 A reagent composition including HMPA instead of TMEDA was used by: Lin, J., Nikaido, M M , Clark, G *J Org Chem* **1987**, *52*, 3745 For other similar reagents for conjugate addition of the vinyl group see Horiguchi, Y , Matsuzawa, S , Nakamura, E , Kuwajima, I *Tetrahedron Lett* **1986**, *27*, 4025, Johnson, C R , Marren, T J *Tetrahedron Lett* **1987**, *28*, 27
- 26 Kende, A. S , Johnson, S , Sanfilippo, P , Hodges, J. D , Jungheim, L N *J Am Chem Soc* **1986**, *108*, 3513
- 27 Pettersson, L Ph D Thesis, Lund University, Sweden, 1989
- 28 Tsunoda, T , Suzuki, M , Noyori, R *Tetrahedron lett* **1980**, *21*, 1357
- 29 For a review on aldolreactions see: Mukaiyama, T *Org React* **1982**, *Vol 28*, 203-331
- 30 Findley, T W , Swern, D , Scanlan, J T *J Amer Chem Soc* **1945**, *67*, 412
- 31 Rubottom, G M , Gruber, J M *J Org Chem* **1977**, *42*, 1051
- 32 Hassner, A , Alexanian, V *Tetrahedron Lett* **1978**, 4475
- 33 Migashita, N , Yoshikoshi, A , Grieco, P A *J Org Chem* **1977**, *42*, 3772
- 34 Corey, E J , Venkateswarlu, A *J Amer Chem Soc* **1972**, *94*, 6190